Long-term effects of juvenile NMDAr or DAr antagonism on adolescent reward-related neurobehavioral outcomes

Kate Goheen

A thesis submitted to the Faculty of Graduate and Postdoctoral Affairs in partial fulfillment of
the requirement for the degree of

Master of Science

in

Neuroscience

Carleton University

Ottawa, Ontario

November 2018

©Kate Goheen
Long-term term effects of juvenile treatment of NMDAr or DAr antagonism on adolescent reward-related neurobehavioral outcomes

Previous research from our laboratory found a critical period of development between postnatal day (P) 18-P24 in which synaptogenesis occurs with the emergence of spatial performance. Considering the glutamatergic projections from the hippocampus to the nucleus accumbens (NAc), it is likely that they share similar developmental trajectories. Subjects were treated with the dopamine receptor (DAr) antagonist, flupenthixol or the NMDAr antagonist, MK-801 from P18-P24 and were tested in an operant conditioning procedure during adolescence. Another group of subjects were given the same drugs prior to each acquisition session to test immediate effects. Spine densities for the early treatment groups were quantified to measure structural changes in the NAc and c-Fos labeling were quantified after an DA or NMDA agonist to measure receptor desensitization. Early flupenthixol increased locomotor activity during acquisition, which corresponded with an increase in DAr sensitization in the NAc. No behavioural or structural differences were found between the early MK-801 group and saline control. Late flupenthixol decreased operant acquisition and locomotor activity, while late MK-801 increased both. These results demonstrate the importance of increased DA in reward-related behaviour, either through the immediate effects of a drug or through DAr sensitization during development.
Acknowledgements

I would like to thank my thesis supervisor Dr. Matthew Holahan for your guidance, ingenuity and wit who made the writing of my thesis both possible and enjoyable. I have learned so much in the past two years, thank you for taking on a squeamish Psychology student.

I would like to thank my committee members Dr. Natalina Salmosa and Dr. Hongyu Sun for taking the time to participate in both my prospectus and defense, as well as thoughtfully providing constructive criticism. I would also like to thank Dr. Maria DeRosa for taking time out of her schedule to be the internal-examiner at my defense.

To my lab-mates/friends for moving buildings with me four times and always staying upbeat, thank you for making the past two years a positive adventure. A special thank you to Erin Noye Tuplin who taught me (sometimes twice) lab techniques and gave me critical advice on surviving graduate school.

I would like to thank the subjects, the rats, without whose death would leave many scientific discoveries unturned. You do not go unnoticed.

Lastly I would like to thank my friends and family who supported me throughout the past two years. Thank you for always listening to me and encouraging me. I owe you one.
Table of contents

Abstract .......................................................................................................................... ii
Acknowledgements ...................................................................................................... iii
Table of Contents ....................................................................................................... iv
List of Figures ............................................................................................................. vi
Abbreviations ........................................................................................................... vii

Introduction ................................................................................................................. 1
  Early History of Memory .......................................................................................... 1
  Biological Basis of Memory .................................................................................... 2
  The Study of Learning & Memory .......................................................................... 4
    Classical & Operant Conditioning ....................................................................... 4
  The role of the Nucleus Accumbens in associative learning .................................. 5
  Anatomy ................................................................................................................ 6
    Afferents ............................................................................................................ 6
    Efferents ......................................................................................................... 6
    Neuron Types ................................................................................................. 7
  Function ................................................................................................................ 8
  Mechanisms of Learning & Memory ..................................................................... 8
  LTP in the Mesolimbic Pathway: glutamate & NMDA ........................................ 11
  LTP in the Mesolimbic Pathway: dopamine .......................................................... 13
  Development of Associative Learning .................................................................... 15
  Purpose ............................................................................................................... 18

Materials and Methods ............................................................................................ 19
  Subjects ............................................................................................................... 19
  Drugs .................................................................................................................. 19
  Behaviour .......................................................................................................... 20
    Drug Treatment ............................................................................................... 20
    Food Restriction .............................................................................................. 20
  Operant Acquisition ............................................................................................. 20
  Statistical Analysis ............................................................................................... 21
  Neural ............................................................................................................... 21
    Drug Treatment ............................................................................................... 21
  Immunohistochemistry ......................................................................................... 21
  Stereology .......................................................................................................... 22
  Golgi Cox Staining .............................................................................................. 23
  Morphological Analysis ....................................................................................... 23

Results ................................................................................................................... 25
Operant Acquisition.................................................................25
Early Treatment.................................................................25
Late Treatment.................................................................30
C-Fos quantification.............................................................36
Dopamine........................................................................36
NMDA..............................................................................39
Spine Density ....................................................................41
Comparison of Total Activity and c-Fos quantification..............43
Discussion.......................................................................44
Summary........................................................................44
Dopamine........................................................................45
NMDA..............................................................................50
Conclusion & Future Directions..............................................58
References.......................................................................59
List of Figures

Figure 1a  Daily Active Lever Presses (Early Treatment)
Figure 1b  Daily Inactive Lever Presses (Early Treatment)
Figure 1c  Daily Port Entries (Early Treatment)
Figure 1d  Daily Locomotor Activity (Early Treatment)
Figure 2a  Daily Active Lever Presses (Late Treatment)
Figure 2b  Daily Inactive Lever Presses (Late Treatment)
Figure 2c  Daily Port Entries (Late Treatment)
Figure 2d  Daily Locomotor Activity (Late Treatment)
Figure 3  Representative Images of c-Fos labeling (Dopamine)
Figure 4a  Mean Cell Density of c-Fos in the NAc core (Dopamine)
Figure 4b  Mean Cell Density of c-Fos in the NAc shell (Dopamine)
Figure 5  Representative Images of c-Fos labeling (Dopamine)
Figure 6a  Mean Cell Density of c-Fos in the NAc core (Dopamine)
Figure 6b  Mean Cell Density of c-Fos in the NAc shell (Dopamine)
Figure 7  Representative Images of Dendritic Spines in the NAc
Figure 8a Total Spine Density in the NAc
Figure 8b  Total Spine Subtype Densities in the NAc
Figure 9  Total Active Lever Presses and Locomotor Activity Compared with c-Fos results
Abbreviations

ABC Avidin-Biotin Complex

AP Action Potential

AP-5 (2R)-amino-5-phosphonopentanoate

APV (2R)-amino-5-phosphonovaleric acid

ANOVA Analysis of Variance

AMPA α-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid

Ca^{2+} Calcium

CaMKII Ca^{2+}/calmodulin-dependent protein kinase II

CNQX Cyanquixaline (6-cyano-7-nitroquinoxaline-2,3-dione)

DA Dopamine

DAB 3,3'-diaminobenzidine

DAr Dopamine receptor

EPSP Excitatory post synaptic potentials

ERK Extracellular signal-regulated kinase

GABA Gamma-aminobutyric acid

LTD Long-term depression

LTP Long-term potentiation

NMDA N-methyl-D-aspartate

NMDAr N-methyl-D-aspartate receptor

Na^+ Sodium

NAc Nucleus Accumbens
Mg+ Magnesium
MK-801
MSN Medium Spiny Neurons
PBS Phosphate buffer solution
PFC Prefrontal Cortex
PKA Protein kinase
P Postnatal day
S-R Stimulus-Response
TZG (RS)-(Tetrazol-5-yl) glycine
VTA Ventral Tegmental Area
SN Substantia Nigra
Introduction

Early History of Memory

The search for the cellular basis of learning and memory is rooted in an epistemological debate between empiricists and rationalists. At the core are fundamental questions pertaining to the study of knowledge, perception and the material basis of the mind; questions that have been documented since the Ancient Greeks (Aristotle, 384-322 BC; Plato, 427-347 BC). Learning was and continues to be, broadly defined as the modification of future behaviour in response to present environmental conditions. Memory can be defined as the ability of an organism to store and retrieve information. Due to insufficient knowledge about the brain, the better part of the philosophical and scientific endeavor into the material basis of learning and memory was rooted in introspective and logical approaches. In the 17th and 18th century rationalists, such as Descartes, concluded that the mind was immaterial (Descartes, 1637), whereas most empiricists abstained from commenting on the nature of the mind (Locke, 1693; Hume, 1748). They focused instead on sensory stimuli, learned associations and habit formation, work that would later influence the founder of experimental psychology, William James (Gluck, Mercado & Myers, 2016). A pursuit originating with good intentions ended with psychologists like Freud and Jung who relied on introspection rather than experimental observation to explain human action and motivation (Gluck, Mercado & Myers, 2016).

In stark opposition to this approach, Watson formed behaviourism, which became the dominant view during the first half of the 20th century (Petri & Mishkin, 1994). Being a firm proponent of empiricism, Watson stated that discussion of consciousness and the mind had no place in the field of psychology (Watson, 1913). Advocates for this school of thought stressed that the only scientifically sound method of studying psychology was through behavioural
observation and viewed learning as a simple stimulus-response association. Cognitive psychologists, from an opposing school of thought, disagreed with such a reductionist view, stating that mental events and expectations contributed significantly to the study of learning (Petri & Mishkin, 1994). The field became stagnant, with behaviourists and cognitivists arguing between the superiority of behavioural observation versus the necessity to include mental events. Throughout this debate, the cellular basis of learning and memory and the emerging field of neuroscience was largely ignored, despite the fact that both could help settle the debate.

**Biological Basis of Memory**

Significant progress in the field of neuroscience occurred when, in 1873, Camillo Golgi developed a method for staining cells that captured axonal and dendritic visibility (Climino, 1999). This led him to support the reticular theory, originally proposed by Joseph von Gerlach, and erroneously conclude that cells in the nervous system are continuous; an idea he continued to espouse despite increasing evidence in favour of nerve cell discontinuity (Cajal 1888, Golgi, 1906; Climino, 1999; Sotelo, 2003). The now established neuron doctrine was originally suggested by Ramon y Cajal when he demonstrated that nerve cells were separate from each other but interconnected by synapses (Cajal, 1906). In addition, he was among the first to propose a theory about the cellular basis of learning (Miles et al., 2005) postulating that as learning occurred, pre-existing connections between neurons strengthened, thereby increasing synaptic efficiency.

Donald Hebb later expanded on the position of Cajal in his book The Organization of Behaviour (Hebb, 1949). The goal was to settle the debate between behaviourists and cognitivists by developing a coherent theory of the mind that redefined it as a physiological entity. In an effort to bridge the gap between psychologists, physiologists, and chemists, he...
believed that it was necessary to merge physiological data from the nervous system with
behavioural data from experimental observation. Examining how they influenced one another
rather than studying them independently would result in a more complete understanding of
human nature. By redefining the concept of the mind as neural activity, he provided a framework
in which researchers from different fields and schools of thought could have dialogue and
collaborate. Hebb’s framework gave behaviourists the opportunity to move away from a simple
stimulus-response explanation of behaviour while keeping their empirical rigor. It permitted
cognitivists to discuss mental processes without sounding “mystical” (Hebb, 1949) and allowed
the narrow focus of physiologists to be broadened by integrating their data with those collected
by psychologists.

An important contribution that Hebb made to the beginnings of the physiological
psychology movement centered on how behavior could change the brain. Hebb proposed that
after repeated stimulation, different “cell-assemblies”, namely a particular set of neurons, would
develop and fire together, often resulting in a permanent change in behaviour. Cell assemblies
only develop when the cells are mutually active and a cell-assembly could occur in response to
external stimuli, activity in a previous cell-assembly or from both. Thoughts were also proposed
to arise from a series of cell assemblies, which Hebb termed a “phase sequence”. Hebb proposed
that cell assemblies could disassemble when repeated stimulation from a particular stimulus was
no longer present. Therefore, from Hebb’s perspective, learning and memory were thought to
occur through the creation and strengthening of synapses, while forgetting occurred through the
weakening of synapse. Hebbian plasticity was theoretically in place, but empirical support was
needed.
The Study of Learning and Memory

Classical & Operant Conditioning

Though radical behaviourism disappeared, many of the procedures put forth by the school of thought are still widely used today. While Watson was viewed as the founder of behaviourism, the beginning of the experimental study of learning can be accredited to the physiologist Ivan Pavlov (Pavlov, 1927). Interested in the physiology of salivation and digestion, Pavlov began to systematically study the salivation of dogs in response to food. He soon discovered that he could elicit a salivary response by a stimulus that predicted the delivery of food. This response became stronger after repeated pairings of a bell with the food and could be elicited by the bell alone. This type of learning in which a previously neutral stimulus (bell) elicits a conditioned response (salivation) after repeated pairings with an unconditioned stimulus (food) is known as classical conditioning. Pavlov also demonstrated a process called extinction, in which a conditioned response is weakened when the conditioned stimulus (bell) is no longer paired with, and thus no longer predicts, the unconditioned stimulus.

During this time, another form of associative learning was discovered in the laboratory of Edward Thorndike (Thorndike, 1911). Instrumental or operant conditioning occurs when a frequency of a behaviour is either increased or decreased following the presence or absence of an outcome. Thorndike observed that cats who were locked in a box were able to be trained to solve a type of puzzle that unlocked the box, specifically their frequency of “unlocking” behaviour increased in order to obtain a favourable outcome. Instrumental, unlike classical conditioning, requires an organism to make a response in order to obtain a specific outcome. Associative learning occurs in all mammals, including humans, making classical and operant conditioning
procedures robust measures of learning that allow experimenters to infer underlying mechanisms across species.

The foundation of behaviourism was largely influenced by the principles of associative learning discovered by Pavlov and Thorndike. The widely used Skinner box, named after the inventor, was developed in 1930 (Gluck, Mercado & Myers, 2016). The original apparatus was a box with a lever that, when pressed, delivered food. An animal placed in the box would explore and eventually press the lever and receive food. Once animals learned the relationship between pressing the lever (response) and the delivery of food (outcome), their rate of response would increase. Since the inception of the Skinner box, there have been many experimental manipulations of the apparatus, allowing researchers to explore the intricacies of operant conditioning and associative learning. For example, standard operant chambers use two levers to assess stimulus discrimination, where pressing one lever results in food delivery, and pressing the other lever does not (Tuplin, Stocco, Holahan, 2015).

During associative learning, two parallel processes occur: both the response-outcome relationship and the value of the reward is learned. When analyzing behavioural data, it is often difficult to distinguish between learning and motivational processes. These two components are likely mediated by different neurochemical pathways, and understanding the subtle nuance between the two may require different behavioural and neurochemical manipulations.

**The role of the Nucleus Accumbens in associative learning**

On a neural level, associative learning occurs through the strengthening of connections between motor neurons (response) and either sensory (stimulus) or cortical neurons (reward expectancy) (Donahoe & Palmer, 1994; Donahoe, Palmer & Burgos, 1997). One pathway involved is the mesocorticolimbic dopamine system, where dopaminergic projections are
activated from the substantia nigra (SN) and ventral tegmental area (VTA) to the dorsal and ventral striatum (Gerfen, Herkenham, & Thibault, 1987).

**Anatomy**

**Afferents**

The nucleus accumbens (NAc) is located in the ventral striatum and can be anatomically divided into a central core, which is surrounded medially, ventrally and laterally by the shell (Zaborsky et al., 1985; Zahm & Brog, 1992). The NAc receives direct dopaminergic input from the VTA and SN, predominantly to the shell and core respectively (Gipson, Kupchick & Kalivas, 2014; Moore & Bloom, 1978; Gerfen, Herkenham & Thibault, 1987; Nirenberg et al., 1996). A subset of the dopaminergic neurons from the VTA express the vesicular transporter VGLUT2 and corelease glutamate in the NAc (Kawano et al., 2006; Yamaguchi et al., 2011). Additionally, independent glutamatergic neurons project to the NAc (Yamaguchi, Sheen & Morales, 2007; Nair-Roberts et al., 2008) and synapse with DA neurons locally (Dobi et al., 2010). The NAc receives direct glutamatergic input from the basolateral amygdala, ventral hippocampus, thalamus, VTA, prelimbic and prefrontal cortex (Gipson, Kupchick & Kalivas, 2014; Powell & Leman, 1976; Kelley et al., 1982). The core predominantly receives input from the dorsal prelimbic, anterior insular, anterior cingulate and perirhinal cortices while the shell receives input from the dorsal peduncular, infralimbic and piriform cortices (Brog et al., 1993; Salgado & Kaplitt, 2014).

**Efferents**

Inhibitory gamma-aminobutyric acid (GABA) pathways project from the NAc to areas in the basal ganglia and midbrain including the ventral pallidum, thalamus and lateral hypothalamus (William, Crossman & Slater, 1977; Churchill & Kalivas, 1994; Floresco, 2015;
The NAc is primarily made up of GABAergic medium spiny neurons (MSNs) expressing D1 and D2 dopamine receptors (Kawaguchi et al., 1995; Yager et al., 2015), the remaining estimated 5% are GABAergic and cholinergic interneurons (Meredith, 1999). MSNs expressing D1 receptors project directly to the SN, whereas MSNs expressing D2 receptors project indirectly to the SN through the globus pallidus (Gerfen, 1992). Though few in comparison, the interneurons are critically involved in the function of the NAc (Kreitzer, 2009). The core is denser than the shell, with larger cell bodies, and a higher density of dendrites and dendritic spines (Meredith, Blank & Groenewegen, 1989; Meredith et al., 1992). The core has more GABAA receptors (Churchill et al., 1992), while the shell has more serotonin receptors (Patel, 1995).

Function

The NAc is a part of the mesocorticolimbic dopamine system, which as a whole, composes the reward system. The NAc itself acts as an interface between motor and limbic systems due to its efferent projections to the former and afferent projections from the latter (Mogenson, Jones & Yim, 1980), making it a prime candidate for initiating behaviour in response to sensory stimuli (Shiflett & Balleine, 2011). It is involved in reward-associative learning, habit formation and compulsive behaviour.

Lesions to the NAc impair instrumental lever pressing for both food and sucrose (Balleine & Kilcross, 1994; Borchgrave et al., 2002). Lesioned rats were not sensitive to changes
in the value of the reward produced by increased food deprivation or sucrose concentration. A
functional dissociation between the NAc core and shell was found (Corbit, Muir, & Balleine,
2001), with NAc core lesioned rats pressing significantly less during acquisition than NAc shell
or sham controls. NAc core lesioned rats were not affected by outcome devaluation, in that they
pressed equally for two rewards that had different values. Lesions to the NAc do not appear to
affect extinction (Wang et al., 2008; Stern & Passingham, 1996), though the NAc core is
involved in reactivation of reward-seeking behaviour after extinction (Wang et al., 2008).

Mechanisms of Learning and Memory

Lomo & Bliss (1973) are credited with the discovery of long-term potentiation (LTP) and
depression (LTD) on the hippocampal synapse between the perforant path and dentate area.
Their seminal work laid the necessary groundwork providing the opportunity for subsequent
researchers to provide empirical evidence in support of Hebbian plasticity. They demonstrated,
first in vitro (Lomo, 1966) and then in vivo that high-frequency presynaptic stimulation gives
rise to long-lasting excitatory postsynaptic potentials (EPSP). The recorded LTP at the
postsynaptic cell outlasted the stimulation from minutes to hours and was the first demonstration
that an event as transient as an action potential could create long-lasting changes. Lomo & Bliss
posited that LTP could be involved in memory storage, partly because it was demonstrated in the
hippocampus, a region viewed as being associated with learning and memory (Lomo & Bliss,
1973; Douglas, 1967). However, they could only hypothesize as to what was occurring and
whether this could occur naturally in an organism. They suggested, among other propositions,
that this increase in synaptic efficiency could be due to an increase in neurotransmitter release
presynaptically or an increase in post-synaptic sensitivity.
Collingridge et al. (1983) sought to examine these hypotheses by experimenting with the CA3-CA1 synapse in the hippocampus. At the time, it was known that this synapse was excitatory, but it was unclear whether the neurotransmitter released was glutamate or l-asparate. Furthermore, the receptor subtypes were just beginning to be categorized and potent antagonists were being created (Davies et al., 1981). With the development of an NMDA receptor (NMDAr) antagonist, Collingridge was able to demonstrate that NMDAr were unnecessary for normal transmission, but that blocking NMDAr prevented LTP induction despite normal depolarization of the post synaptic target. Concurrently, it was demonstrated that intracellular injections of a calcium blocker in CA1 neurons blocked LTP induction indicating that LTP was dependent on calcium influx (Lynch et al., 1983; Miles et al., 2005). Both results supported the notion that LTP expression occurred at the post and not the presynaptic membrane. Kullman (1994) was able to confirm this hypothesis by recording receptor specific synaptic signals at baseline compared to after LTP induction. Through this, he discovered that AMPA receptor signals change but NMDAr do not, indicating that the former is latent at baseline and increase after LTP. These findings lead him to propose that a rise in AMPA receptors was behind the expression of increased synaptic efficiency. Therefore, the same amount of presynaptic neurotransmitter release would have more receptors to bind to, causing a larger depolarization.

It is now known that LTP and LTD occur at excitatory glutamatergic synapses, where the ionotropic AMPA receptor (AMPAr) allows the influx of sodium (Na+) ions, but the voltage-dependent NMDA receptor channel is blocked by magnesium (Mg+) (Dingleidine et al., 1999). With enough presynaptic glutamatergic stimulation, Na+ floods in through AMPAR channels and pushes out the magnesium (Mg+) block through electrostatic repulsion allowing Na+ and calcium (Ca2+) influx via NMDAR. The level of postsynaptic Ca2+ determines whether LTP or
LTD occur (Lisman, 1989). In LTP, Ca2+ binds to calmodulin (CaM) and activates the Ca2+/calmodulin-dependent protein kinase II (CaMKII) (Malenka et al., 1988) which can facilitate the insertion of AMPAr into the post-synaptic neuron (Zamanillo et al., 1999). In LTD, less Ca2+ enters the postsynaptic cell, causing an activation of protein phosphatases, eventually leading to AMPAr internalization (Beattie et al., 2000).

Hebb, in his theory of plasticity, proposed 1) an increase in synaptic efficiency when neurons fire mutually and 2) the creation of connections between neurons, presently referred to as synaptogenesis. The first part of the theory was verified through the finding that synaptic strengthening or weakening was expressed through levels of AMPAr postsynaptically as well as the finding that NMDAr act as coincident detectors, requiring both pre and postsynaptic stimulation for LTP induction (Zorumski & Izumi, 2012). The second part of the theory requires a process expanding further than the synapse.

Much of what is known of LTP and LTD is from work on the hippocampus. However, LTP and LTD are more than the cellular basis of learning and memory, rather they can be thought of as a form of synaptic plasticity. Neural plasticity occurs in other regions besides the hippocampus, such as the mesolimbic pathway (Bonci & Malenka, 1999; Robbe, Bockaert & Manzoni, 2002) and many genes related to plasticity including those that encode protein kinases and cAMP response element-binding protein (CREB) are found in the striatum and cortex (Kelley, 2004). The study of neural plasticity covers a much broader range of topics including development, post-injury, addiction, epilepsy, and pain.

**LTP in Mesolimbic pathways: Glutamate and NMDA**

The role of dopamine in reward-based learning has been the primary focus of research but the role of glutamate is of equal importance. As stated previously, the NAc receives
excitatory glutamatergic input from the prelimbic and prefrontal cortex, as well as the basolateral amygdala, hippocampus, and VTA. The induction of LTP and LTD has been demonstrated in the VTA (Bonci & Malenka, 1999; Bellone & Lusher, 2005), amygdala (Nabavi et al., 2014) and NAc (Robbe, Bockaert & Manzoni, 2002; Meredith, Pennartz & Groenewegen 1993). Most research on synaptic plasticity in the striatum has focused on corticostriatal synapses, rather than limbic-striatal synapses (Cerovic et al., 2013).

Calabresi et al. (1992) conducted one of the first studies demonstrating synaptic plasticity at corticostriatal synapses. EPSPs were induced by tetanic stimulation of cortical inputs, and this was blocked using the AMPAr antagonist cyanquixaline (6-cyano-7-nitroquinoxaline-2,3-dione) (CNQX), but not by the NMDAr antagonist (2R)-amino-5-phosphonovaleric acid (APV). When slices were bathed in a Mg2+ control condition, tetanic cortical stimulation produced LTD, demonstrating that tetanic stimulation in regular conditions produces LTD rather than LTP. In a Mg2+ free medium, cortical stimulation produced LTP, which was then blocked by an AMPAr antagonist, but not an NMDAr antagonist, demonstrating an NMDAr independent form of LTP.

It is now understood that striatal LTP varies and can be influenced by numerous factors including region, the age of development, and whether the experiment is in vivo or in vitro (Filippo et al., 2009). LTP in the NAc has been evoked using high-frequency stimulation, but this method is not ecologically valid because high frequency (100 Hz) LTP is found in less than 20% of MSNs (Kombian & Malenka, 1994; Schramm, Egli & Winder., 2002) and most MSNs fire between 1 and 10 Hz (Carelli & Ijames, 2000). Glutamatergic neurons in the prefrontal cortex (PFC) and amygdala that project to the NAc also fire at a similar rate (Margrie, Brecht & Sakmann, 2002; Puig et al, 2003) Therefore, using a spike-timing-dependent plasticity model, a method that pairs action potential with EPSPs at a lower frequency may be more physiologically
relevant than high-frequency stimulation when studying synaptic plasticity in the NAc (Ji & Martin, 2012).

Ji & Martin (2012) investigated synaptic plasticity in MSNs in the NAc core using varying conditions by combining different action potential (AP) frequencies (0.2, 1 and 5 Hz) with varying delays between AP and ESPS pairings (10, 20, 50 and 200ms). Two subgroups of MSNs emerged across experimental conditions, in which either LTP or LTD was induced. LTP-MSNs were influenced by the frequency and delay of AP-EPSPs, whereas LTD-MSNs were not. An NMDA antagonist blocked LTP-MSNs, whereas antagonizing calcium channels blocked LTD-MSNs. These data underscore the challenging nature of studying synaptic plasticity in the NAc.

The behavioural effect of NMDAr blockade was demonstrated using an operant task (Kelley, Smith-Roe, & Holahan, 1997). Rats were trained to lever press for a food reward and received infusions of an NMDAR antagonist (2R)-amino-5-phosphonopentanoate (AP-5) before or after training. NMDAR dependent plasticity in the NAc core, basolateral amygdala, and medial PFC was found to be necessary for the acquisition of the task. Infusions of AP-5 in the NAc shell impaired learning to a lesser degree. Once the task was learned, AP-5 had no effect indicating that NMDA is necessary during the early stages of learning but not the expression phase. The necessity of synaptic plasticity for instrumental learning was further demonstrated by inhibiting protein synthesis following acquisition trials. Synaptic plasticity in the NAc core, but not the shell, was necessary for memory consolidation during acquisition. Once the task was learned, protein synthesis inhibition had no effect (Hernandez, Sadeghian & Kelley, 2002).
**LTP in the Mesolimbic Pathway: dopamine**

MSN in the striatum contain both dopaminergic and glutamatergic receptors located in close proximity to each other (Smith & Bolam, 1990; Sesack et al., 2003; Shiflett & Balleine, 2011). The NAc receives dopaminergic input from the VTA and SN, both synapsing on striatal MSNs which express either D1 or D2 dopaminergic receptors (Smith & Bolam, 1990; Shiflett & Balleine, 2011). Excitatory glutamatergic afferents are thought to interact with DA neurons from VTA to NAc, increasing repetition of behaviours under certain environmental conditions (Chen, Hopf & Bonci, 2010).

DA released from midbrain neurons act as a signal to predict reward, by firing when there is a discrepancy between reward expectation and behavioural outcome (Schultz, 1999). For example, in Pavlovian conditioning, the VTA fires and DA is released to unexpected rewards. However, once the delivery of a reward is predicted, VTA neurons no longer fire. Instead, they fire to the presentation of the conditioned stimulus, that is, they fire to the prediction of a reward. During extinction, DA firing is decreased when the reward is no longer presented, signaling that the conditioned stimulus no longer predicts the reward.

In a study using intracranial self-stimulation of the SN, rats lever pressed for electrical stimulation, which released DA onto MSNs (Reynolds, Hyland & Wickens, 2001). After the task was learned, the rate at which each rat pressed the lever was calculated. Electrical stimulation was applied to the SN at the individualized rate for each rat producing synaptic potentiation at corticostriatal synapses. The degree of potentiation induced by the optimal electrical stimulation was correlated to the rate at which each rat learned, suggesting that DA helps strengthen corticostriatal synapses and is associated with the rate of instrumental learning. Potentiation was
blocked using a D1 receptor antagonist, suggesting that DA contributes to corticostriatal synaptic plasticity through these receptors.

LTP has frequently been observed in MSNs expressing D1 receptors, (Calabresi et al., 2000; Kerr & Wickens, 2001). However, in the absence of D1 receptor binding, LTD occurs when these corticostriatal synapses are activated (Shen et al., 2008). This suggests that DA acts as a signal, strengthening the synapses when the cortical environmental input and striatal behavioural output is rewarding. Importantly, phasic DA firing and not single-bursts of DA is necessary for LTP induction. This is because D1 receptors have low affinity to DA and require high concentrations for binding (Creese et al., 1983; Gonan, 1997). In the absence or low levels of DA, LTD occurs, reducing the likelihood that the same cortical input will result in striatal output. For example, if a rat is given the choice of two levers and is expecting the delivery of a food pellet, pressing the correct lever will result in DA firing and a strengthening between the cortical reward expectancy and the striatal action (Shiflett & Balleine, 2011; Horvitz, 2009). Pressing the incorrect lever will result in LTD in that active corticostriatal synapse, decreasing the likelihood of future incorrect lever presses.

Intracellular molecules that act as coincidence detectors to DA and glutamate are critically involved in synaptic plasticity and instrumental behaviour (Shiflett & Balleine, 2011; Nakano et al., 2010). For example, extracellular signal-regulated kinase (ERK) reaches maximum activity following the combined activity of D1 and glutamate receptor activation (Girault et al., 2007). Many proteins involved in synaptic plasticity, such as protein kinase, CREB, and c-Fos are regulated by DAr and NMDAr (Kelley, 2004). D1 and D2 receptors act in opposing ways on intracellular pathways, partly by increasing or decreasing adenylyl cyclase, respectively (Greengard, 2001; Surmeier et al., 2007). An increase in adenylyl cyclase increases
cAMP and cAMP-dependent protein kinase (PKA). PKA strengthens synaptic plasticity by phosphorylating ion channels, receptors, and transcription factors, downstream effects result in an increase in postsynaptic AMPAr insertion (Surmeier et al., 2007). D2 receptor activation decreases adenylyl cyclase and reduces cellular excitability (Surmeier et al., 2007). The subtypes of MSNs demonstrated by Ji & Martin (2012) were likely D1 and D2 expressing MSNs.

**Development of Associative Learning**

Processes that govern synaptic plasticity, such as LTP and LTD, also govern the development of neural circuits (Durand, Kovalchuk & Konnert, 1996; Ben-Ari et al., 1997). During the first week of postnatal development in the hippocampus, glutamatergic synapses gradually become functional (Durand, Kovalchuk & Konnert, 1996). During P1 and P2 only NMDA receptors exist and by P6, they are colocalized with AMPAr. A pairing protocol was used in slices of P1 hippocampal neurons in which presynaptic stimulation was paired with postsynaptic depotentiation. The natural development of glutamatergic synapse was reproduced and the purely NMDA synapses became functional. This “synaptic induction” was blocked by the AMPA antagonist CNQX, indicating that the development and function of glutamatergic receptors was due to the presence of AMPAr (Goodman & Shatz, 1993; Choi & Lovinger, 1997).

The processes involved in synaptic plasticity and development occur in tandem, often influencing one another. The theory that the environment can influence the trajectory of neural development and that these changes can influence subsequent plasticity is referred to as metaplasticity, or the plasticity of plasticity (Kania et al., 2017). First coined by Abraham & Bear (1996), metaplasticity refers to the idea that when studying plasticity, it is not only important to consider what is happening at present during LTP and LTD, but to acknowledge the history of
the neuron and synapse as well, which may influence the capacity of plasticity at that particular synapse.

Developmental metaplasticity has been mainly studied in the visual cortex, but information gained from these studies is likely relevant to regions involved in associative learning. For example, Kirkwood, Rioult & Bear, (1996) demonstrated that during a critical period of visual development, the threshold at which presynaptic glutamatergic stimulation triggers either LTP or LTD can change based on the environment, in this case, light deprivation. Importantly, this effect can be reversed after light exposure during the critical period. This threshold modification is likely due to changes in NMDAr subunits during development.

NMDAr consist of four subunits: two NR1 and two that can be NR2 (A, B, C or D) or NR3 (A or B) (Cull-Candy & Leszkiewics, 2001). The combination of subunits determine the function (Cull-Candy & Leszkiewics, 2004). The ratio of NR2A to NR2B changes during development, and it is hypothesized that this may affect LTP and LTD by changing the permeability of NMDAr to Ca2+ (Yashiro & Philpot, 2008). However light deprivation may disrupt the normal developmental ratio of NR2A/NR2B, thereby disrupting the threshold at which LTP and LTD occur. It is reasonable to believe that similar processes may be disrupted by exposure to environmental factors during different critical periods during the development of other neural regions.

Unlike sensory and motor neural regions, regions underlying associative learning have a longer developmental trajectory. Previous research has determined a sensitive period in hippocampal development between P18-P24 in which synaptogenesis correlates with the emergence of spatial performance (Tzakis et al., 2016). Considering the glutamatergic
projections from the hippocampus to the NAc and PFC, they likely share a similar developmental period (Zaczek, Hedreen & Coyle 1979; Jay & Witter 1991).

The mesolimbic dopamine system of the rat continues to develop throughout the first four postnatal weeks (Voorn et al., 1988; Tepper, Trent & Nakamura, 1990; Wang & Pitts, 1994). In the nucleus accumbens, DA neurons continue to distribute and grow past PN week three (Voorn et al., 1988). Nigrostriatal dopamine neurons do not reach maximum velocity until PN week three and adult firing patterns until PN week four (Tepper, Trent & Nakamura, 1990). In the prefrontal cortex, the density of DA neurons continues to increase until P60 (Kalsbeek et al., 1988). The properties of striatal neurons change throughout development (Choi & Lovinger, 1997). When comparing striatal slices from P10-P19 with P23-P27, the likelihood of neurotransmitter release as well as the magnitude of LTD induction decreased with age.

Rodent adolescence occurs from approximately P28-P42 (Spear, 2000), but may extend to P55 which would coincide with late adolescents and emerging adulthood seen in humans (Vetter-O’Hagen & Spear, 2012). Adolescent humans and laboratory animals are particularly sensitive to rewarding stimuli (Doremus-Fitzwater et al., 2010; van Duijvenvoorde et al., 2016), are more sensitive to palatable foods (Friemel, Spanagel, & Schneider, 2010) and exhibit increased goal-directed behaviour (see review Doremus-Fitzwater & Spear, 2016).

Purpose

The purpose of this thesis was to determine the contribution of NMDAr and DAr in the development of associative learning circuitry by using an instrumental conditioning assay. This was done by using chronic treatment with either the NMDAr antagonist MK-801 or the DAr antagonist flupenthixol during a critical period of development (P18-24) followed by instrumental conditioning in adolescence. Two hypotheses were explored to help explain what
form of developmental plasticity contributed to the behavioural outcomes. The structural plasticity hypothesis posited that early treatment with either an NMDAr antagonist or DAr antagonist would disrupt the structural integrity of the NAc, manifesting as reduced spine density. The synaptic plasticity hypothesis posited that preadolescent DAr or NMDAr antagonism would desensitize receptors in adolescence. Receptor desensitization would manifest by reduced c-Fos labelling following an acute injection of the DA agonist apomorphine or the NMDA agonist TZG. These neural measures would help explore plasticity both structurally and at the level of the synapse. Looking at both dendritic spine density as well as receptor function allows us to gain insight into how the neurons in the NAc might be responding during operant conditioning.
Materials and Methods

Subjects

Pregnant Long-Evans (n=15) female rats were obtained from Charles River, Quebec and were single-housed in clear plastic cages (25 x 20 x 45 cm). They were given ad libitum access to food and water and were housed in a temperature controlled environment with a 12-hour light/dark cycle (lights on at 8 am). The day the pups were born was recorded as postnatal day (P) 0. Female pups were culled at P12 and the males were weaned at P18. Pups were housed in groups of 3 or 4, were given a red plastic tube for enrichment and had ad libitum access to food and water. Pups were either used for behaviour (n= 48) or neural analysis (n=30). Procedures were approved by the Carleton University Animal Care Committee and were in agreement with the Canadian Council on Animal Care.

Drugs

The noncompetitive NMDAr antagonist (+)-5-methyl-10, 11-dihydro-5H-dibenzo [a, d] cyclohepten-5-10-imine maleate (MK-801) (Sigma-Aldrich) was dissolved in 0.9% sterile saline to the concentration of 0.05mg/kg with dH2O and stored frozen. MK-801 at doses of 0.05mg/kg, 0.1mg/kg and 0.2mg/kg reached maximal concentration an hour post injection (IP) (Wegener et al., 2011). The 0.05mg/kg reached a maximal concentration of 6.02nM, while doses of 0.1mg/kg and 0.2mg/kg reached concentrations of 14.34nM and 34nM, respectively. Because MK-801 has been demonstrated to occupy receptors at a concentration of 3nM (Reynolds, Murphy & Miller, 1987), the dose chosen for the present work would be sufficient to produce NMDAr blockade. The nonselective DA antagonist Flupenthixol (Sigma-Aldrich) was diluted to a concentration of 0.25mg/kg. 0.9% sterile saline was used as a control injection. The NMDA agonist (RS)-(Tetrazol-5-yl) glycine (TZG) (Sigma-Aldrich) is a highly selective and potent NMDA agonist.
that readily crosses the blood brain barrier (Schoepp et al., 1991). TZG was diluted to a sub-seizure concentration of 0.1247mg/kg. The nonselective DA agonist Apomorphine (Sigma-Aldrich) was diluted to the concentration of 3mg/kg.

**Behaviour**

**Drug Treatment**

From P18-P24 half of the pups (n=24) used for behaviour were injected subcutaneously each day with either saline, MK-801 or Flupenthixol at a volume of 0.1ml. Rats that did not receive an early treatment with a drug were randomly assigned to receive subcutaneous injections of MK-801, flupenthixol or saline 15 minutes prior to the beginning of each trial at a volume of 0.1ml.

**Food Restriction**

One week prior to, and during operant conditioning (around P30) pups (n=48) were single housed and food restricted to approximately 90% of their weight using an aged matched control.

**Operant Acquisition**

Six conditioning chambers (Coulborn Instruments, Whitehall, PA; 30.5 cm x 25.5 cm x 30.5 cm) were used to conduct the operant conditioning procedures. Chambers were made of Plexiglas and stainless steel and were located in ventilated, sound attenuating boxes. Two levers were located on the right wall with a port in between. A house light signaled the beginning of each session. During acquisition, rats were placed into the chambers for 30 minutes for a total of five days. Every uninterrupted second response (FR2 schedule) on the left lever (active lever) resulted in the delivery of a 45 mg chocolate pellet (BioServe, New Jersey). This was paired with
the cue light located above the left lever switching from red to green and the house lights extinguishing. Presses on the right lever (inactive) were not rewarded.

**Statistical analysis**

The acquisition (and extinction) data were analyzed using a two-way, repeated measures analysis of variance (ANOVA) with treatment group as the between factor. For the acquisition data total active lever presses, inactive lever presses, port entries or locomotor activity over the 5 days were used as the repeated measure. For extinction, data after each five-minute interval was used as the repeated measure. Post-hoc comparisons were performed if ANOVAs were significant.

**Neural Analysis**

**Drug Treatment**

The second group of pups (n=30) were treated with MK-801, flupenthixol or saline at a volume of 0.1 ml daily from P18-P24. From P33-P41, they were single caged and food restricted to 90% of their weight. A P41 they were given an injection of TZG, apomorphine or saline. Rats pretreated with MK-801 received an injection of TZG or saline (IP, 0.3 ml). Rats pretreated with flupenthixol received an injection of apomorphine or saline (IP, 0.3 ml). Rats pretreated with saline received an injection of saline, TZG or apomorphine. 90 minutes later, rats were perfused and their brains were removed.

**Immunohistochemistry**

One hemisphere from each subject was fixed in 4% paraformaldehyde in 0.01 M phosphate buffer solution (PBS, pH 7.4). The following day they were placed in a 30% sucrose in 0.1 M PBS and stored at 4°C. Brains were sectioned on a Leica CM1900 cryostat (Weitzler,
Germany) at 35 um and placed in well plates filled with a 0.1% sodium azide solution in 0.1 M PBS.

Sections were washed three times for 7 minutes in 0.01M PBS solution with 0.2% Triton X (PBS-TX). They were incubated in hydrogen peroxide (H\textsubscript{2}O\textsubscript{2}) PBS-TX (1:100) for 15 minutes followed by 3, 5 minute washes in PBS-TX. Sections were then blocked in a 1x animal free blocker (AFB; Vector) in PBS-TX for 30 minutes and were transferred to the primary antibody (rabbit anti c-Fos, 1:5000) for two nights at room temperature. The tissue was then washed 3 times for 10 minutes in PBSTX, and incubated in the secondary antibody (anti-rabbit, 1:1000). Sectioned were washed 3 times for 10 minutes in PBSTX and then incubated in Avidin-Biotin Complex (ABC) for 1 hour. They were then washed 3 times for 5 minutes in 0.01M PBS and developed in 3,3'-diaminobenzidine (DAB) until exposed. Sectioned were washed 2 times for 5 minutes in PBS, mounted, dehydrated and cover slipped.

**Stereology**

The number of c-Fos labelled cells in the NAc core and shell were estimated using unbiased stereological principles and the optical fractionator method. Sections were visualized using an Olympus BX51 bright field microscope with a motorized stage (Olympus Canada, Markham, ON). An Olympus U-CMAD3 camera was used to capture images. Stereo Investigator (MBF Bioscience, Williston, VT) software was used for unbiased stereological quantification. A magnification of 2.5x was used to trace the NAc core and shell, and a 60x magnification lens (oil immersion, NA 1.35) was used to count the c-Fos positive cells. Two sections per region were used per rat, with a minimum of 3 rats per group. The estimated number of c-Fos cells using weighted section thickness was calculated through Stereo Investigator. One data point per subject was calculated by taking the average cell count from each section. In order
to produce a standardized data point across subjects, the mean estimated number of c-Fos cells was divided by the area (mm$^2$). Data were analyzed using a one-way ANOVA, post-hoc tests were performed if significant group differences were found.

**Golgi Cox Staining**

The other hemisphere from subjects were fixed for seven days in a potassium dichromate, mercuric chloride and potassium chromate solution (Golgi fix solution). Brains were washed in dH$_2$O for 4 hours, 3 hours, overnight and then cryoprotected in 10% sucrose for 8 hours, 20% sucrose overnight and 30% sucrose for a minimum of 4 days.

Using a Vibratome, the brains were sectioned at 200 μm, placed on gelatinized slides, and stored in a dark, humidified box for 24 hours. Sections were washed with DH$_2$O for 1 minute, immersed in 28% Ammonium Hydroxide for 40 minutes and rinsed in DH$_2$O for 1 minute. They were then submerged in Kodak film fix A (diluted 1:1 with DH$_2$O) for 40 minutes, and rinsed in DH$_2$O for 1 minute two times. They were then immersed in 50%, 70% and 95% ethanol for 1 minute each. The solutions from this point on were desiccated with type 3A molecular sieve, 1/16” pellets. The slides were immersed three times in 100% ethanol for 5 minutes, and then immersed in 33% ethanol, 33% Clearene and 33% chloroform solution for 10 minutes. They were then submerged in a Clearene solution for 30 minutes. Slides were then cover slipped with Permount mounting medium and placed in a desiccated box for a minimum of 4 days before analysis.

**Morphological analysis**

Neurolucida software (MBF Bioscience Inc., Williston, VT) was used to trace MSN dendrites at 100x magnification using an Olympus BX51 microscope. A total of 3 neurons from the NAc were selected for each rat. Only rats pretreated with MK-801, flupenthixol or saline and
given an acute injection of saline were used for analysis. Three dendrites per neuron were
selected, each with a minimum length of 200 mm. Spine subtypes have been classified as stubby,
mushroom, thin, filopodia (Peters & Kaiserman-Abramof, 1970; Garcia-Lopez, Garcia-Marín &
Freire, 2006) and can be differentiated based on shape (Jedynak et al., 2007; Harris et al., 1992).
Mushroom and stubby spines have a head diameter > 0.055 μm, filopodia and thin spines have a
head diameter < 0.055 μm. Mushroom and filopodia spines have a discernible neck, where as
thin and stubby spines do not. Spine density was calculated by averaging the total spine count
divided by the total length of each dendrite per neuron. Data were analyzed using a one-way
ANOVA, post-hoc tests were performed if significant group differences were found.
Results

Operant Acquisition

Two-way repeated measures ANOVAs were conducted on all dependent variables using day as the within factor and treatment as the between factor. Early and late treatment groups were analyzed separately.

Early Treatment

Analysis of the mean number of active lever presses per day (Figure 1a) revealed a main effect of day \([F(4,116) = 33.65, p < .000]\), but no main effect of treatment \([F(2,29) = 2.062, p=0.145]\) and no day x treatment interaction \([F(8,116) = 1.394, p=.206]\). Analysis of the mean number of inactive lever presses per day (Figure 1b) revealed a main effect of day \([F(4,116)= 4.988, p <0.001]\), but no main effect of treatment \([F(2,29)= 1.574, p=0.224]\) and no day x treatment interaction \([F(8,116)= 20.717, p=0.672]\).

Analysis of the mean number of port entries per day (Figure 1c) revealed a main effect of day \([F(4,116) = 13.207, p<0.001]\), but no main effect of treatment \([F(2,29) = 2.343, p =0.114]\) and no day x treatment interaction \([F(8,116)= .918, p =0.504]\). Analysis of mean locomotor activity per day (Figure 1d) revealed no main effect of day \([F(4,116) = 1.195, p =0.317]\), but a main effect of treatment \([F(2,29) = 11.428, p <0.001]\) and a significant day x treatment interaction \([F(8,116) = 3.329, p < 0.01]\). Tukey post-hoc tests on the effect of treatment revealed that group pretreated with flupenthixol showed significantly higher locomotor activity than the group treated with saline \((p < 0.01)\) and the group treated with MK-801 \((p < 0.001)\).
Figure 1a. Total active lever presses for chocolate pellets (FR2) during 30 minute acquisition sessions (mean ± SEM). No group differences were found.
Figure 1b. Inactive lever presses during acquisition sessions (mean ± SEM). No group differences were found.
Figure 1c. Total port entries during acquisition sessions (mean ± SEM). No group differences were found.
Figure 1d. Total locomotor activity during acquisition (mean ± SEM). The group pretreated with flupenthixol showed significantly higher levels of locomotor activity compared to both the saline (p < 0.01) and the MK-801 (p < 0.001) groups.
Late Treatment

Analysis of mean number of active lever presses per day (Figure 2a) revealed a main effect of day \( [F(4,112) = 27.942, \ p < 0.001] \), a main effect of treatment \( [F(2,28) = 9.058, \ p < 0.001] \) and a significant day x treatment interaction \( [F(8,112) = 6.711, \ p < 0.001] \). Tukey post-hoc tests on the treatment effect revealed the group treated with MK-801 pressed the active lever significantly more than the group treated with flupenthixol \( (p < 0.001) \).

Follow up one-way ANOVAs on each day were conducted. A one-way ANOVA on day 3 revealed a main effect of treatment \( [F(2,30)=6.786, \ p < 0.01] \). Tukey post-hoc tests revealed that the group treated with MK-801 pressed the active lever significantly more than groups treated with saline \( (p < 0.05) \) and the group treated with flupenthixol \( (p < 0.01) \). On day 4, a one-way ANOVA revealed a main effect of treatment \( [F(2,30)=6.434, \ p < 0.01] \). Tukey post-hoc tests revealed that the group treated with flupenthixol pressed significantly less than the group treated with saline \( (p < 0.05) \) or MK-801 \( (p < 0.01) \). On day 5, a main treatment effect was found \( [F(2,30) = 12.622, \ p < 0.001] \), with the flupenthixol group showing less presses than the groups treated with saline \( (p < 0.05) \) or MK-801 \( (p < 0.01) \).

Analysis of the mean number of inactive lever presses per day (Figure 2b) revealed a main effect of day \( [F(4,112) = 3.942, \ p < 0.01] \), but no main effect of treatment \( [F(2,28) = 3.106, \ p = 0.060] \) and no day x treatment interaction \( [F(8,112) = 0.548, \ p = 0.818] \).

Analysis on mean number of port entries per day revealed a main effect of day \( [F(4,112)=9.926, \ p < 0.001] \), a main effect of treatment \( [F(2,28)=10.581, \ p < 0.01] \), and a day x treatment interaction \( [F(8,112)=2.821, \ p < 0.01] \). Tukey post-hoc tests revealed that the group treated with MK-801 had significantly more port entries than the group treated with flupenthixol \( (p < 0.01) \). Analysis of locomotor activity per day revealed a main effect of day \( [F(4,112)=] \).
2.558, p < 0.05], a main effect of treatment \([F(2, 28) = 20.253, p < 0.01]\) and a day x treatment interaction \([F(8,112) = 2.47, p < 0.05]\). Tukey post-hoc tests revealed that the group treated with flupenthixol showed significantly less locomotor activity than the groups treated with saline or MK-801 (p < 0.001).
Figure 2a. Total active lever presses for chocolate pellets (FR2) during acquisition sessions (30 minutes each) (mean ± SEM). On day 3, the group that received an injection of MK-801, 15 minutes prior to each acquisition session pressed the active lever significantly more than the groups treated with saline (p < 0.05) or flupenthixol (p < 0.01). On day 4 and 5, the group treated with flupenthixol pressed significantly less than the group treated with saline (p < 0.05) or MK-801 (p < 0.01).
Figure 2b. Inactive lever presses during acquisition sessions (mean ± SEM). No group differences were reported.
Figure 2c. Total port entries during acquisition (mean ± SEM). The group treated with MK-801, 15 minutes prior to each acquisition session showed significantly more port entries than the group treated with flupenthixol (p < 0.01).
Figure 2d. Locomotor activity during acquisition (mean ± SEM). The group injected with flupenthixol 15 minutes prior to each acquisition session showed significantly less locomotor activity than the groups treated with saline or MK-801 (p < 0.001).

**C-Fos quantification**

**Dopamine**

The average cellular density [mean cell count/ volume (µm³)] for each treatment group was calculated. Data are represented as percent change from control [(average cellular density of individually treated rats/ group average of pretreated and acutely injected with saline) * 100]. Rats were pretreated with saline or flupenthixol during preadolescence and received an injection
of apomorphine or saline at test. A one-way ANOVA on the density of c-Fos labeling in the NAc core (Figure 4a) revealed a main effect of treatment $[F(2,12)= 9.114, p < 0.01]$. Tukey post-hoc tests revealed a higher density of c-Fos labeling in the group pretreated with flupenthixol and injected with apomorphine at test compared to the group pretreated with flupenthixol and treated with saline at test ($p < 0.05$).

A one-way ANOVA on the density of c-Fos labeling in the NAc shell (Figure 4b) revealed a main effect of treatment $[F(2,12)= 4.243, p < 0.05]$. Tukey post-hoc tests revealed a higher density of c-Fos labeled cells in the group pretreated with flupenthixol and injected with apomorphine at test compared to the group pretreated with flupenthixol and treated with saline at test ($p < 0.05$).
Saline-Apo

Flu-Apo

Figure 3. Representative images of c-Fos for the dopamine treatment groups in the NAc (60x)
Figure 4a & b. Percent change from control of the average cellular density [mean cell count/volume (µm$^3$)] of c-Fos labeling in the core and shell (mean ± SEM). The group pretreated with flupenthixol that received an acute injection of apomorphine at test showed significantly more c-Fos labeling in the core and the shell than the group pretreated with flupenthixol that received an acute injection of saline ($p < 0.05$).

NMDSA

Rats were pretreated with saline or MK-801 during preadolescence and received an injection of TZG or saline at test. A one way ANOVA on the density of c-Fos labeling in the NAc core (Figure 5a) and shell (Figure 5b) revealed no main effects of group [$F(2,12)= 3.650$, $p = 0.065$]; [$F(2,12) = .650$, $p = 0.543$].
MK-801-Saline

Saline-TZG

MK-801-TZG

Figure 5. Representative images of c-Fos for the NMDA treatment groups in the NAc (60x)
Figure 6a & b. Percent change from control of the average cellular density [mean cell count/volume (µm$^3$)] of c-Fos labeling in the core and shell (mean ± SEM). No group differences were found.
**Spine density quantification**

A one-way ANOVA on total spine density (Figure 5a) revealed no main effect of treatment \( [F(2,6) = 0.643] \). A two-way repeated measures ANOVA on spine density subtype (Figure 6b) revealed a main effect of spine subtype \( [F(3,35) = 23.407, p < 0.01] \), but no main effect of treatment \( [F(2,35) = 0.765] \). Tukey post hoc tests revealed significantly more thin spines than stubby, mushroom and filopodia \( (p < 0.01) \) and more stubby spines than mushroom and filopodia \( (p < 0.01) \).

![Representative images of dendritic spines in the NAc at 100x.](image-url)

**Saline**

**MK-801**

**Flupenthixol**
Figure 8a & b. Total spine density [total number of spines/total length (µm)] and spine subtypes [number of each spine subtype/total length (µm)] in the NAc (mean ± SEM). A main effect of spine subtype was found (p < 0.01), but no main effect of treatment.
Discussion

Summary

Rats were treated with MK-801, flupenthixol or saline during a preadolescent period (P18-24) and were tested in an operant conditioning procedure after adolescence. Early treatment with the dopamine receptor antagonist, flupenthixol, increased locomotor activity and lever pressing during the post-adolescent testing phase (Fig 9). Early treatment with the NMDA receptor antagonist, MK-801, did not result in any overt behavioral alterations during the test. To examine the behavioral changes under the influence of the drug, separate groups of rats were treated with flupenthixol or MK-801 prior to each acquisition session. In this case, flupenthixol (0.25 mg/kg; same dose as used during preadolescent phase) decreased lever pressing and locomotor activity (Fig 9). While early MK-801 had no effect on any behavioral measure, late MK-801 was associated with an increase in bar pressing, port entries and locomotor activity (Fig 9).

Immunohistochemical and morphological analyses were conducted to provide a potential explanation of the behavioural results. Two possible hypotheses were explored to assess what form of developmental plasticity contributed to the behavioural outcomes. The structural plasticity hypothesis stated that early treatment with either an NMDA receptor antagonist or DA receptor antagonist would impede the development of neurons in the NAc and this would manifest as reduced spine densities. The synaptic plasticity hypothesis stated that early treatment with MK-801 or flupenthixol would desensitize NMDA or DA receptors, respectively, during adolescence. This would manifest as reduced c-Fos labeling following an injection of the NMDA agonist TZG or the dopamine receptor agonist apomorphine. There was no evidence of structural differences in the NAc between treatment groups, possibly indicating that behavioural
differences seen were not due to morphological changes. Elevated c-Fos staining was found after an acute challenge with the dopamine agonist apomorphine in both the saline-apo and flupenthixol-apo group. This increase in cellular activity was higher in the flupenthixol group compared to controls, indicating a sensitized response at DA receptors. It is likely that the elevated bar pressing and locomotor activity as seen in the early flupenthixol group was due to sensitized DA receptors in the NAc (Fig 9). The NMDA agonist, TZG, slightly elevated cellular activity in both the saline-TZG and MK-801-TZG group. Interestingly basal c-Fos levels were equal in the MK-801-saline group compared to TZG-induced levels.

Comparing total active lever presses and total locomotor activity with the c-Fos results (Figure 9) we can see that early flupenthixol increased activity and apomorphine-induced c-Fos. This provides support for a dopaminergic sensitization hypothesis whereby preadolescent treatment with a DA antagonist sensitized dopaminergic receptors in the NAc and elevated operant conditioning in adolescence.

![Graph showing total presses over 5 days](image)

![Graph showing total locomotor activity over 5 days](image)
lever presses and total locomotor activity over 5 days of acquisition with c-Fos results after apomorphine.

**Dopamine**

Early flupenthixol (0.25mg/kg) increased lever pressing and activity during the adolescent test (Fig. 9), whereas flupenthixol treatment during the adolescent test (0.25mg/kg) decreased both. One possible explanation for the divergent treatment effects is that early treatment with flupenthixol sensitized DAr. During operant conditioning, DA in the NAc is first released in response to the reward then shifts to the cue predictive of the reward (Schultz, Dayan & Montague, 1997; Day et al., 2007). Once the cue becomes conditioned and after presentation of a CS, mesolimbic DA neurons activate (Waelti, Dickinson & Schultz, 2001) and release DA into the NAc (Cheng, Bruin & Feenstra, 2003; Day et al., 2007; Stuber et al., 2008). Likewise, NAc DAr blockade impairs instrumental response to a CS (Wakabayashi, Fields & Nicola 2004; Nicola, 2010). Given that extracellular DA in the NAc increases locomotor activity (Pijnenburg & van Rossum, 1973) and lever pressing (Salamone et al., 1990; Wyvell & Berridge, 2000), chronic early treatment with flupenthixol may have sensitized DAr in the NAc. This would increase the efficacy of the CS-induced DA release (Sandstrom, Nelson & Bruno, 2003); either due to an increase in the number of postsynaptic DAr receptors or an increase in the sensitivity of DAr present (Nielsen et al., 1973)

We tested the hypothesis that preadolescent flupenthixol treatment altered DAr sensitivity by challenging subjects with an acute injection of the DA agonist apomorphine during adolescence (P40). This day was chosen because it was equivalent to the first day of operant acquisition, allowing us to infer what may be happening at the level of the synapse during the operant task. The early flupenthixol and the early saline groups were given an acute injection of
apomorphine (3mg/kg, s.c). This dose was chosen because it increases locomotor activity (Castro et al., 1985; Fredriksson & Archer, 2000) and it is a threshold dose in between 2mg/kg (Cole et al., 1992) and 4mg/kg (Wirtshafter, 2000) that increase c-Fos levels in the striatum. While low (1mg/kg) (Rebec et al., 1979; Deutch & Dunman, 1996) and high doses (5mg/kg) (Pennypacker, Zhang & Hong, 1992; Filipkowski, Rydz & Kaczmarek., 2001; Saint Marie et al., 2006) resulted in inconsistent levels of c-Fos (Filipkowski, Rydz & Kaczmarek., 2001), the medium dose consistently increases c-Fos.

Elevated apomorphine-induced c-Fos levels were found in the NAc in both the flupenthixol-apo and saline-apo group. These results are consistent with previous research demonstrating increased locomotor activity and NAc activation following apomorphine (Steiner & Gerfen, 1994; Cenci et al., 1992) or other DA agonists (Kalivas & Duffy, 1990; Johnson & Glick, 1993; Scofield et al., 2016). This increase in cellular activity was higher in the early-flupenthixol treated group compared to the early-saline treated control, indicating potential dopaminergic sensitization in the early flupenthixol group. These results corroborate the behavioural results that revealed increased locomotor activity and elevated lever presses in the early-flupenthixol group and support the hypothesis that increased locomotor activity and total active lever presses were due, in part, to sensitized dopaminergic receptors in the NAc.

In a previous study, rats were pretreated with flupenthixol (2.5mg/kg, orally) for 12 days and tolerance towards the pharmacological properties of the DAr antagonist were tested following various time periods of withdrawal (Nielsen et al., 1974). On the test day, pretreated rats and controls were first given an acute injection of flupenthixol followed by apomorphine. Flupenthixol pretreated rats exhibited tolerance to apomorphine; higher doses of acute flupenthixol were needed to block apomorphine-induced stereotypy. This effect was present for
up to 21 days following withdrawal, and highest 3 days after. In the present study, apomorphine
induced c-Fos levels were higher in the flupenthixol group compared to saline group.
Considering that the time period of withdrawal in the present study was 7 days, it is likely that
similar synaptic processes occurred.

After acute DAr blockade and withdrawal, there is increased sensitivity to apomorphine
(Nielsen et al., 1973). Studies have shown that after long-term treatment with the antipsychotic
and D2 receptor antagonist haloperidol, rats and humans show increased locomotor activity and
elevated densities of D2 receptors (see Review Seeman et al., 2006). Indeed, patients with
schizophrenia who have taken antipsychotics have been shown through post-mortem studies
(Seeman, 1987) and PET scans (Nordstrom et al., 1995; Tune et al., 1993) to have increased D2
receptor density. Chronic treatment with flupenthixol has been demonstrated to increase D2
receptor density (Hess, Norman & Creese, 1988). Flupenthixol in vitro has an equal affinity for
D1 and D2 receptors, but in vivo preferentially interacts with D2 receptors which may explain
why D2, but not D1 receptor density increased (Hess, Norman & Creese, 1988). Co-
administration of both D1 and D2 receptor antagonists increased apomorphine-induced
locomotion and D2 receptor density (Braun, Laruelle & Mouradian, 1996).

Morphological analysis revealed no differences in structural changes in the NAc between
the early saline and early flupenthixol groups. This suggests that preadolescent DAr blockade
does not produce long-term structural changes in the NAc. In another study preadolescent rats
(P27) were administered neurotoxin 6-hydroxydopamine (6-OHDA) in the medial forebrain
bundle resulting in depletions of DA in striatum (Sandstrom, Nelson & Bruno, 2003). A week
post-surgery locomotor activity and basal DA levels in the NAc were measured. Compared to
aged-matched controls, lesioned rats had similar locomotor activity and DA levels despite having
a 95% reduction in DA producing cells. Adults rats that were lesioned showed an equal depletion in DA producing cells one week post 6-OHDA administration, yet exhibited severe motor deficits as well as an 80% reduction in basal DA levels. Based on these results and our current study there seem to be compensatory mechanisms during preadolescence following DAr blockade or lesion.

As previously stated, extracellular DA in the NAc is required for operant acquisition, therefore blocking DAr through receptor occupation would impair this process. As expected, late treatment with flupenthixol (0.25mg/kg) decreased locomotor activity and total lever pressing. This, along with other research, shows that the immediate effect of dopamine receptor blockade is to reduce locomotor activity and bar pressing (Floresco, Tse & Ghods-Sharifi, 2008). The same dose (0.25mg/kg) decreased the frequency of high effort responding (FR10) but did not affect the latency towards the lever (Floresco, Tse & Ghods-Sharifi, 2008). A lower dose (0.125mg/kg) did not have immediate effects on locomotor activity yet decreased bar pressing (Floresco, Tse & Ghods-Sharifi, 2008) suggesting that DAr blockade impairs lever pressing ability independently from the motoric impairing effects. Similarly, the lower dose (0.125mg/kg) did not decrease lever pressing performance when an instrumental task was already learned (Dickinson, Smith & Mirenowicz, 2000). When there is choice between a large reward with a delay and a small immediate reward, 0.5mg/kg, but not 0.25mg/kg effects the likelihood of subjects choosing the latter (Wade, Wit & Richards, 2000).

Another dopamine antagonist, Pimozide, affects operant conditioning in a dose-dependent manner, whereby a lower dose decreased acquisition independently from locomotor activity or lever pressing ability (Yokel & Wise 1976; Wise & Schwartz, 1981). Similarly, both a D1 and D2 receptor antagonist decreased lever pressing, but not food consumption (Salamone et
al., 2002). Others have found that the D2 receptor antagonist haloperidol suppressed lever pressing for food (Salmone et al., 1991) and decreased high effort choices for a large reward in a T-maze (Denk et al., 2005). In a Pavlovian procedure in which a stimulus (lever) is followed by a reward (sucrose) it has been observed that some subjects will find the lever itself rewarding (sign-trackers) while some subjects will go to the port to wait for the reward (goal-trackers). Low doses of haloperidol disrupt sign-tracking, but not goal-tracking, demonstrating that DAR antagonism may disrupt the process by which a cue or CS becomes rewarding (Danna & Elmer, 2010).

In operant conditioning, port entries are a behavioural measure used to directly assess reward-seeking behaviour and indirectly assess frequency of reward received. In the present study, late flupenthixol treatment (0.25mg/kg) decreased port entries. However, it is difficult to discern whether the low port entries were due to an inability to enter the port or due to lack of reward-seeking behaviour. A higher dose of flupenthixol (0.5mg/kg) did not impair port entries when a reward was present (Wade, Wit & Richards, 2000), therefore it is probable that the lower dose (0.25mg/kg) in the current study did not impair the ability to enter the port. Rather, the decrease in port entries was likely due to less reward present and not ability to obtain the reward.

Taken together, these results demonstrate and highlight two points: when studying the effects of an antagonist, it is 1) difficult to distinguish whether the blockade interferes with the learning process itself or interferes with a behaviour required to demonstrate learning (Beninger, 1983) and 2) the dose and subtype of the receptor help dissociate learning processes from motor ability. It is evident that a varying degree of DAR occupation is needed for certain aspects of operant acquisition and performance. A wider dose range in the current study may have helped dissociate locomotor activity from lever pressing and port entries.
In the present study, early MK-801 (0.05mg/kg) did not produce a learning deficit; no group differences were found in active lever pressing, inactive lever pressing or port entries during acquisition between the early MK-801 group and saline control. Late MK-801, on the other hand (0.05mg/kg), enhanced acquisition and increased total port entries. The early treatment results are surprising given the importance of NMDAr in developmental plasticity (Durand, Kovalchuk & Konnert, 1996; Ben-Ari et al., 1997) and associative learning (Chen, Hopf & Bonci, 2010; Shiflett & Balleine, 2011). We hypothesized that early NMDAr antagonism would decrease both structural and synaptic plasticity in the NAc and consequently impair operant learning. Morphological analysis revealed no structural changes in the NAc between early-MK-801 and saline groups. This is consistent with the behavioural results that revealed no differences during operant conditioning.

The goal of the agonist test during adolescence was to examine the influence of NMDAr antagonism during preadolescence on later receptor function. The NMDA agonist TZG slightly elevated c-Fos expression in the NAc in both the saline-TZG and MK-801-TZG groups, potentially indicating increased cellular activity compared to control in response to ligand binding. Previous research found TZG (Rogers et al., 2005; Inada et al., 2007) and NMDA increased c-Fos expression (Sonnenberg et al., 1989; Lerea, Butler & McNamara, 1992) and increased Ca2+ (Lerea, Butler & McNamara, 1992) in neural regions including the NAc (Inada et al., 2007; Dragunow & Faull, 1990; Duncan et al., 1998). No differences were found between rats pretreated with MK-801 and saline in response to TZG which is consistent with the
behavioural data that demonstrated similar lever pressing, port entries and locomotor activity in both groups.

Interestingly, rats pretreated with MK-801 and injected with saline had the highest levels of c-Fos labeling. This is consistent with previous research demonstrating increased c-Fos levels in the hippocampus after repeated injections with the NMDAr antagonist ketamine followed by a 14 day washout period (Keilhoff et al., 2004). One possible explanation for these results is a supersensitization of NMDAr where LTP is more easily induced. Therefore, neutral stimuli or a saline injection may produce increased c-Fos levels that would not normally occur due to a lowering of the threshold needed for activation (Keilhoff et al., 2004).

During development, NMDAr subunits change with an increase in NR2A subunit expression (Monyer et al., 1994; see review Yashiro & Philpot, 2008). In rats, the ratio of NMDAr containing NR2B change from 70% at P-15-P28 to 40% of adult levels at P31-P49 (Wang & Gao, 2009). It is hypothesized that this change may affect LTP and LTD by changing the permeability of NMDAr to Ca$^{2+}$ (Yashiro & Philpot, 2008). An increase in the ratio of NR2A/NR2B at a synapse is thought to change the LTP/LTD crossover threshold making it more likely for LTD to occur.

In one study, high doses of systemic MK-801 (1mg/kg, i.p.) were injected in mice from P16 to P40 (Inta et al., 2017). C-Fos levels peaked in the hippocampus at P24 followed by lower levels at P30 and P40. It was hypothesized that MK-801 increased c-Fos levels by binding to NR2B-containing NMDAr which then disinhibited GABAergic interneurons (Gass et al., 1993; Inta et al., 2017). The differences found in MK-801 induced c-Fos expression during development may be due to NMDAr subunit changes. Indeed, following administration of the
NR2B specific antagonist Ro 25-6981, c-Fos levels peaked at P24 and declined thereafter (Inta et al., 2017).

Changes in NMDAr subunits are partly mediated by sensory experience (Yashiro & Philpot, 2008). During a critical period of visual development, light deprivation will influence the switch from NR2B to NR2A to occur more slowly (Nase et al., 1999; Roberts & Ramoa, 1999). It is unclear how NMDAr antagonism in the present study would impact the developmental switch of these glutamatergic receptors. In adulthood, repeated MK-801 increases NR2B subunits in the hippocampus (Rujescu et al., 2006), which increases likelihood of LTP to occur (Barria & Malinow, 2005; Bellone & Lusher, 2012; Mameli et al., 2011). The rise in NR2A is activity dependent; glutamate is required to bind to NMDAr for protein synthesis to occur (Barria & Malinow, 2005). If glutamate binding to NMDAr is necessary for NR2A production, it is possible that antagonizing these receptors may slow down this process. If early MK-801 did slow down this process, there would be a higher ratio of NR2B receptors during the agonist test compared to the saline control and an increased likelihood of LTP to occur. This may explain the NMDAr supersensitivity and increased basal c-Fos levels seen after an injection of saline (Keilhoff et al., 2004). Future studies would benefit from using western blot analyses to measure levels of protein for NMDAr subunits and compare the early-MK-801 treatment with saline controls during adolescence.

When comparing the present study to the literature, differences in behavioural outcomes after repeated MK-801 administration can be characterized in to differences in timing, dose and behavioural task. To our knowledge NMDAr blockade during this period of development (P18-P24) has not been tested in an operant procedure. Previous research has demonstrated that rats pretreated with MK-801 (0.1mg/kg, twice daily) from P7-P11 exhibited severe learning deficits.
during operant acquisition in adulthood (Rapanelli et al., 2013). This is contrary to the present
findings where early MK-801 (P18-P24) did not disrupt acquisition. Rapanelli et al., (2013) also
demonstrated that rats treated with MK-801 (0.1mg/kg twice a day IP) in adulthood one week
prior to operant conditioning exhibited mild learning deficits. The differences between this and
the present study are likely due to both the timing and dose of the injections.

Morphological analysis revealed no structural changes in the NAc between the MK-801
and saline groups. Novak et al. (2013) characterized 60 genes during striatal development and
found that during P7-P14 an early gene network that does not express striatal-specific genes was
down-regulated and replaced with a mature network. This network gave immature neurons a
functional identity and projection pathway. During this week, there was a rise in D1 and D2
receptors, both following the same developmental trajectory. At the end of this week MSNs
found their functional identity. Repeated injections with MK-801 from P7-P11 during the time
where neurons are developing their functional identity would likely have a severe impact on
striatal development and associative learning. Novak et al., (2013) also found a rise in
myelination genes from P14 to P17. It is possible that NMDAr antagonism after myelination
(P18-P24) had less or no impact on operant conditioning compared to disrupting this
developmental process at an earlier stage.

A meta-analysis of postnatal treatment with MK-801 revealed conflicting results in a
number of behavioural tasks (Lim, Taylor & Malone, 2012). Early NMDAr antagonism does not
seem to effect novel object recognition in adulthood (Delacour, 1988; Stefani & Moghaddam,
2005; Lim, Taylor & Malone, 2012) or visual-cue performance in the Morris water maze (Gorter
& de Bruin., 1992; Nemeth et al., 2002). Postnatal MK-801 treatment impaired spatial
recognition in adult rats (Gorter & de Bruin., 1992; Nemeth et al., 2002; Su et al., 2011), but not
in adolescent rats (Su et al., 2011). Spatial tasks may be more affected because MK-801 has a higher affinity to bind in the hippocampus compared to the cerebral cortex and striatum (Wong et al., 1986).

MK-801 (0.05mg/kg) prior to each acquisition day enhanced acquisition and increased port entries. This is consistent with work from our lab and others that have found low doses of MK-801 (0.025mg/kg and 0.05mg/kg) increased operant acquisition (Sanger & Jackson, 1989; Gilmour et al., 2009; Smith et al., 2011) and lever pressing during extinction (Tuplin, Stocco & Holahan, 2015; Holahan, Clarke & Hines, 2010). In contrast, another study found no effect of low doses (0.025mg/kg and 0.05mg/kg) on operant acquisition (Port, Murphy, Magee & Seybold, 1996). High doses (0.2mg/kg and 0.25mg/kg) consistently impair operant acquisition (Gilmour et al., 2009; Wozniak et al., 1990; Clissold, Ferkany & Pontecorvo, 1991; Pitts et al., 2006; Smith et al., 2011; Lover-Ulecia et al., 2018; Povroznik et al., 2014). There are mixed results with moderate doses (0.1mg/kg) with some finding a mild deficit in learning (Stephani & Moghaddam, 2005; Pitts et al., 2006; Rapanelli et al., 2013; Smith et al., 2011; Lover-Ulecia et al., 2018) and others finding no effect (Clissold, Ferkany & Pontecorvo, 1991; Port, Murphy, Magee & Seybold, 1996). These conflicting results within the same doses of MK-801 are likely due to differences in test parameters (Liu et al., 2017).

The enhancing effect of late MK-801 in the current study may be due to MK-801 working as both an NMDAr antagonist and DA agonist (Hiramatsu, Cho & Nabeshima, 1989; Mathe et al., 1999). A similar NMDAr antagonist and DAr agonist ketamine has been shown to increase LTP in the NAc and hippocampus through D1 receptor activation (Belujon & Grace, 2014). MK-801 has been demonstrated to increase burst firing of dopaminergic VTA neurons (French & Cici, 1990; Belujon & Grace, 2014) and increase DA release while decreasing DA
reuptake in the NAc (Hancock & Stamford, 1999). During acquisition, VTA neurons initially fire to the presentation of a reward and then fire to the presentation of the conditioned stimulus (Schultz, 1999). It is likely that MK-801 primarily acts in the VTA when injected systemically because similar behavioural effects are seen when it is injected directly into the VTA (Narayanan et al., 1996). Therefore, MK-801 may facilitate the formation of stimulus-response associations by increasing NAc DA via the VTA. Similarly, MK-801 (0.1mg/kg) may increase food craving, demonstrated by increased place preference conditioning (PPC) (Yonghui et al., 2006). Furthermore, NMDAr antagonism has been demonstrated to increase extracellular glutamate (Moghaddam et al., 1997; Sebban et al., 2002; Lorrain et al., 2003). Glutamate, unlike DA, can activate or inactive neurons within milliseconds (Lapish, et al., 2006) and may further increase the stimulus-response associations by enhancing dopaminergic signals from the VTA.

In the present study the late MK-801 group (0.05mg/kg) had elevated locomotor activity compared to saline. This is inconsistent with previous research in our laboratory and others that found that a low dose (0.05mg/kg) did not affect locomotor activity (Tuplin, Stocco & Holahan, 2015; Carey, Dai & Gui, 1998; Liljequist, 1991; Nakagawa & Iwasaki, 1996; Gilmour et al., 2009; Liu et al., 2017). However, moderate (0.1mg/kg) (Tuplin, Stocco & Holahan, 2015; Ouagazzal et al., 1994; Narayanan et al., 1996; Kesby et al., 2012; Zavitsanou et al., 2014; Hasegawa et al., 2016; Druhan, Rajabi & Stewart 1996; Hiramatsu, Cho & Nabeshima, 1989; Gilmour et al., 2009; Liu et al., 2017; Rung et al., 2005) and high doses (0.2mg/kg-0.5mg/kg) increased locomotor activity (Tricklebank et al., 1989; Bygrave et al., 2016; Druhan, Rajabi & Stewart 1996; Hiramatsu, Cho & Nabeshima, 1989; Gilmour et al., 2009; Liu et al., 2017; Rung et al., 2005). Increased locomotion is likely due to an increase in DA in the dorsal and ventral striatum (Ouagazzal et al., 1994; Narayanan et al., 1996) as well as the VTA (Narayanan et al., 1996). Similarly, DA antagonists have been demonstrated to reverse
MK-801 induced locomotor activity (Hoffman, 1992; Lapin & Rogawski, 1994; Mele et al., 1996). The rats used in the current study were adolescence therefore the increased locomotion compared to past studies seen after the low dose (0.05mg/kg) could be due to lower body weight.

**Conclusion & Future Directions**

To conclude, early treatment with Flupenthixol enhanced operant conditioning possibly through sensitized DAr in the NAc, demonstrated by increased cellular activity after apomorphine in comparison to the saline control. No morphological changes were found between treatment groups, indicating that behavioural differences seen were not do to structural changes in the NAc. Flupenthixol given before operant acquisition decreased operant conditioning and locomotor activity, likely through DAr blockade (Floresco., Tse & Ghods-Sharifi, 2008).

Early treatment with MK-801 did not produce a learning deficit and did not produce morphological changes in the NAc. However, basal levels of c-Fos were higher in the early MK-801 group, possibly due to NMDAr subunit changes and thus an increased likelihood for LTP to occur (Yashiro & Philpot, 2008). Future studies would benefit from using Western blot analyses to measure protein levels of NR2B and NR2A subunits and compare the early-MK-801 treatment with saline controls during adolescence. MK-801 administrated before operant condition enhanced acquisition and locomotor activity. MK-801 is both an NMDAr antagonist and DA agonist (Hiramatsu, Cho & Nabeshima, 1989) and likely helps form stimulus-response associations through VTA neurons releasing DA into the NAc (Schultz, 1999; French & Cici, 1990).

There are some inconsistencies in the literature regarding both flupenthixol and MK-801 in operant conditioning and development. Future studies may benefit from having a dose-
response curve to better elucidate the role of DA$\alpha_r$ and NMDA$\alpha_r$ in the developmental and immediate effects of both flupenthixol and MK-801 on associative learning. However, the current study discovered important information, as stated above. Taken together, these results demonstrate the importance of increased DA in operant acquisition and locomotion, either through the immediate effects of a drug or through DA$\alpha_r$ sensitization during development.
References


Neuroscience, 12(8), 2973–2981.


Robbe, D., Bockaert, J., & Manzoni, O. J. (2002). Metabotropic glutamate receptor 2/3-dependent long-term depression in the nucleus accumbens is blocked in morphine withdrawn mice. The European Journal of Neuroscience, 16(11), 2231–2235.


for food but increase free food consumption in a novel food choice procedure. *Psychopharmacology, 104*(4), 515–521


Yamaguchi, T., Sheen, W., & Morales, M. (2007). Glutamatergic neurons are present in the rat
ventral tegmental area: Glutamatergic neurons in the VTA. *The European Journal of Neuroscience*, 25(1), 106–118.


